High resolution epigenetics: Direct multiomics simultaneously maps DNA methylation and chromatin proteins

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Gene expression is controlled by crosstalk via **DNA methylation and chromatin proteins**

- > **DNA methylation** (DNAme) & **chromatin proteins** (histone PTMs, transcription factors) coordinate to oversee gene expression
- \succ These factors are associated with specific genomic features (Figure 1)
- > Current methods to study DNAme/protein interactions rely on indirect parallel assays
- > Improved technologies are needed to resolve DNAme / chromatin protein crosstalk



Figure 1. Chromatin proteins and DNAme define genomic features and govern gene expression; however, currently available assays are correlative.

CUT&RUN-EM simultaneously profiles direct

CUT&RUN-EM is highly reproducible across targets, sequencing depths, and DNA inputs



CUT&RUN-EM reveals distinct DNAme profiles at defined genomic features



Figure 6. Representative genome browser tracks (A) demonstrate that CUT&RUN-EM reflects known biological functions of chromatin proteins (e.g. unmethylated H3K4me3



DNAme / chromatin protein interactions



Figure 2. CUT&RUN-EM integrates EpiCypher CUT&RUN with NEB EM-seq v2. CUT&RUN isolates DNA associated with chromatin proteins of interest. EM-seq then enzymatically converts unmethylated cytosines to uracils. Sequencing the resulting libraries enables CpG resolution of DNAme co-occurring with the chromatin mark of interest.

Figure 4. CUT&RUN-EM is highly reproducible across various sequencing depths (A; downsampled genome browser tracks) and replicates (B; Pearson correlation coefficients).



promoters, methylated H3K36me3 gene bodies) when overlaid with DNAme. The association of chromatin proteins with DNAme varies across cell lines (B), highlighting the utility of CUT&RUN-EM to provide cell-specific gene regulatory insights.

CUT&RUN-EM deconvolutes epigenetic crosstalk that is masked in correlative assays



Figure 7. CUT&RUN-EM stratifies subpopulations of cells with unique PTM-DNAme crosstalk signatures that are obscured in whole genome EM-seq (WGEM). IGV genome browser comparison of CUT&RUN (C&R), CUT&RUN-EM (C&R-EM), and WGEM in K562 cells. C&R-EM loci with concordant results (red box) recapitulate the findings of WGEM, while discordant loci (blue box) resolve PTM-DNAme specific signatures.

MeCP2-targeted CUT&RUN (meCUT&RUN) generates ultra-sensitive, global DNA methylation profiles at low cost



Figure 8. (A) Schematic representation of a GST-tagged MeCP2 methyl binding domain used in lieu of an antibody in traditional CUT&RUN) to enrich methylated DNA. Sequencing libraries can be prepared via standard library prep to provide a snapshot of genome-wide DNAme at 150 bp resolution, or unmethylated cytosines can be converted with EM-seq to achieve CpG resolution. (B) MeCUT&RUN (orange) with 500k K562 cells shows high concordance with Methyl-DNA Immunoprecipitation sequencing (MeDIP-seq; purple) at >2.5-fold reduced sequencing depth with 10-fold fewer cells. MeCP2 CUT&RUN-EM (yellow boxed track) generates similar DNA methylation profiles compared to whole genome EM-seq (bottom track). (C) Sankey plot of the binned human genome (200 bp) comparing the presence of 5mC in WGEM and meCUT&RUN-EM. MeCP2 enriches for regions of high 5mC concentration with 34x less sequencing than WGEM, greatly reducing sequencing costs.

Conclusions

- CUT&RUN-EM enables a direct interrogation of DNAme and chromatin protein crosstalk
- MeCUT&RUN provides fast and efficient mapping of global DNA methylation levels with ~150 bp resolution

MeCUT&RUN-EM resolves base pair resolution of DNA methylation with 34x less sequencing than WGEM:

Whole Genome EM-seq		MeCP2 CUT&RUN-EI
\$1,650	VS	\$48

EpiCypher assays in action

- > T-cell activation Arce, Nature 2025 (PMID: <u>39663454</u>)
- T-cell exhaustion Ford, Sci Immunol 2022 (PMID: <u>35930654</u>)
- > T-cell anticancer activity Mamedov, Nature 2024 (PMID: <u>37648854</u>)
- CAR T-cell expansion

(PMID: <u>36944333</u>)

dCas9/Cas9 targeting Wang, Nucleic Acids Res 2022 (PMID: 35849129)

- > Epigenome editing Rohm, Cell Genom 2025 (PMID: <u>39947136</u>)
- > iPSC profiling Schreiber, Mol Metab 2021 (PMID: <u>34352411</u>)
- > Immune cell fingerprinting

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